

## New England Biolabs Certificate of Analysis

**Product Name:** *Bst 2.0 WarmStart<sup>®</sup> DNA Polymerase*  
**Catalog #:** *M0538S/L*  
**Concentration:** *8,000 units/ml*  
**Unit Definition:** *One unit is defined as the amount of enzyme that will incorporate 25 nmol of dNTP into acid insoluble material in 30 minutes at 65°C.*  
**Lot #:** *0101712*  
**Assay Date:** *12/2017*  
**Expiration Date:** *12/2019*  
**Storage Temp:** *-20°C*  
**Storage Conditions:** *10 mM Tris-HCl, 50 mM KCl, 1 mM DTT, 0.1 mM EDTA, 0.1 % Triton<sup>®</sup>X-100, 50 % Glycerol, (pH 7.1 @ 25°C)*  
**Specification Version:** *PS-M0538S/L v1.0*  
**Effective Date:** *21 Nov 2017*

Assay Name/Specification (minimum release criteria)	Lot #0101712
<b>Endonuclease Activity (Nicking)</b> - A 50 µl reaction in ThermoPol <sup>®</sup> Reaction Buffer containing 1 µg of supercoiled PhiX174 DNA and a minimum of 500 units of <i>Bst 2.0</i> DNA Polymerase incubated for 4 hours at 65°C results in <10% conversion to the nicked form as determined by agarose gel electrophoresis.	<b>Pass</b>
<b>Exonuclease Activity (Radioactivity Release)</b> - A 50 µl reaction in ThermoPol <sup>®</sup> Reaction Buffer containing 1 µg of a mixture of single and double-stranded [ <sup>3</sup> H] <i>E. coli</i> DNA and a minimum of 500 units of <i>Bst 2.0</i> DNA Polymerase incubated for 4 hours at 65°C releases <0.1% of the total radioactivity.	<b>Pass</b>
<b>Inhibition of Primer Extension (Hot Start)</b> - A 50 µl reaction in Isothermal Amplification Buffer containing 6 mM MgSO <sub>4</sub> and 1.4 mM dNTPs in the presence of 1.6 µM of a fluorescent internally labeled oligonucleotide and a minimum of 16 units of <i>Bst 2.0</i> WarmStart <sup>®</sup> DNA Polymerase incubated for 2 hours at 25°C yields <5% extension as determined by capillary electrophoresis.	<b>Pass</b>
<b>Non-Specific DNase Activity (16 Hour)</b> - A 50 µl reaction in NEBuffer 2 containing 1 µg of T3 DNA in addition to a reaction containing Lambda-HindIII DNA and a minimum of 120 units of <i>Bst 2.0</i> WarmStart <sup>®</sup> DNA Polymerase incubated for 16 hours at 16°C results in a DNA pattern free of detectable nuclease degradation as determined by agarose gel electrophoresis.	<b>Pass</b>
<b>Phosphatase Activity (pNPP)</b> - A 200 µl reaction in 1M Diethanolamine, pH 9.8, 0.5 mM MgCl <sub>2</sub> containing 2.5 mM <i>p</i> -Nitrophenyl Phosphate (pNPP) and a minimum of 100 units <i>Bst 2.0</i> DNA Polymerase incubated for 4 hours at 37°C yields <0.0001 unit of alkaline phosphatase activity as determined by spectrophotometric analysis.	<b>Pass</b>



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<p><b>Protein Purity Assay (SDS-PAGE)</b> - <i>Bst</i> 2.0 DNA Polymerase is <math>\geq 99\%</math> pure as determined by SDS-PAGE analysis using Coomassie Blue detection.</p>	<b>Pass</b>
<p><b>qPCR DNA Contamination (<i>E. coli</i> Genomic)</b> - A minimum of 120 units of <i>Bst</i> 2.0 WarmStart<sup>®</sup> DNA Polymerase is screened for the presence of <i>E. coli</i> genomic DNA using SYBR<sup>®</sup> Green qPCR with primers specific for the <i>E. coli</i> 16S rRNA locus. Results are quantified using a standard curve generated from purified <i>E. coli</i> genomic DNA. The measured level of <i>E. coli</i> genomic DNA contamination is <math>\leq 1</math> <i>E. coli</i> genome.</p>	<b>Pass</b>
<p><b>RNase Activity (Extended Digestion)</b> - A 10 <math>\mu</math>l reaction in NEBuffer 4 containing 40 ng of a 300 base single-stranded RNA and a minimum of 1 <math>\mu</math>l of <i>Bst</i> 2.0 WarmStart<sup>®</sup> DNA Polymerase is incubated at 37°C. After incubation for 16 hours, <math>&gt;90\%</math> of the substrate RNA remains intact as determined by gel electrophoresis using fluorescent detection.</p>	<b>Pass</b>



Authorized by  
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21 Nov 2017



Inspected by  
Tony Spear-Alfonso  
05 Jan 2018

